

101.811-12 – including *Taq* polymerase, IFU-01
 101.811-12u – without *Taq* polymerase, IFU-02

Visit www.olerup.com for
 “Instructions for Use” (IFU)

Lot No.: **6F3**

Lot-specific information

Olerup SSP[®] DRB4*01:03:01:02N

| | |
|----------------------------------|---|
| Product number: | 101.811-12 – including <i>Taq</i> polymerase 101.811-12u – without <i>Taq</i> polymerase |
| Lot number: | 6F3 |
| Expiry date: | 2020-04-01 |
| Number of tests: | 12 |
| Number of wells per test: | 2+1 |
| Storage - pre-aliquoted primers: | dark at -20°C |
| - PCR Master Mix: | -20°C |
| - Adhesive PCR seals | RT |
| - Product Insert | RT |

This Product Description is only valid for Lot No. 6F3.

Complete product documentation consists of generic Instructions for Use (IFU), lot specific Product Insert, Worksheet and Certificate.

The DRB4*01:03:01:02N specificity and interpretation tables have been updated compared to the previous *Olerup SSP[®] DRB4*01:03:01:02N* lot (**Lot No. 6E4**). The kit design is based on IMGT/HLA database 3.28.0.

The DRB4*01:03:01:02N primer set is unchanged compared to the previous *Olerup SSP[®] DRB4*01:03:01:02N* (**Lot No. 6E4**).

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Well 3 contains Negative Control primer pairs, that will amplify more than 95% of the *Olerup SSP*[®] HLA Class I, DRB, DQB1, DPB1 and DQA1 amplicons as well as all the amplicons generated by the control primer pairs matching the human growth hormone gene.

HLA-specific PCR product sizes range from 75 to 200 base pairs.
The PCR product generated by the positive control primer pair is 430 base pairs.

| Length of PCR product | 105 | 200 | 105 | 80 | 75 | 80 | 85 |
|------------------------------|----------------------|-------------------------|----------------------|----------------------|----------------------|----------------------|----------------------|
| 5'-primer¹ | 164 | 340 | 440 | 45 | 45 | 43 | 36 |
| | 5'-CAC ^{3'} | 5'-Agg ^{3'} | 5'-TTA ^{3'} | 5'-Tgg ^{3'} | 5'-Tgg ^{3'} | 5'-Tgg ^{3'} | 5'-TAC ^{3'} |
| | | | | | | | 36 |
| | | | | | | | 5'-TAT ^{3'} |
| 3'-primer² | 231 | 2nd I | 507 | 59 | 58 | 57 | 47 |
| | 5'-TgC ^{3'} | 5'-AAA ^{3'} | 5'-TTg ^{3'} | 5'-CTC ^{3'} | 5'-ggC ^{3'} | 5'-CTC ^{3'} | 5'-ACA ^{3'} |
| | | | | | | | 48 |
| | | | | | | | 5'-gCA ^{3'} |
| | | | | | | | 48 |
| | | | | | | | 5'-gCC ^{3'} |
| | | | | | | | 52 |
| | | | | | | | 5'-TgT ^{3'} |
| A* | + | + | + | | | | |
| B* | + | + | + | | | | |
| C* | + | + | + | | | | |
| DRB1 | | | | + | + | | |
| DRB3 | | | | + | + | | |
| DRB5 | | | | + | | | |
| DQB1 | | | | | + | | |
| DPB1 | | | | | | + | |
| DQA1 | | | | | | | + |

¹The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon, matching the specificity-determining 3'-end of the primer is given. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

²The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon or the 2nd intron, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

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Lot-specific information

PRODUCT DESCRIPTION

DRB4*01:03:01:02N SSP subtyping

CONTENT

The primer set contains 5'- and 3'-primers for identifying the DRB4*01:03:01:02N allele.

PLATE LAYOUT

Each test consists of 3 PCR reactions in an 8 well cut PCR plate. Wells 4 to 8 are empty.

| | | | | | | | |
|----------|----------|-----------|-------|-------|-------|-------|-------|
| 1 | 2 | NC | empty | empty | empty | empty | empty |
|----------|----------|-----------|-------|-------|-------|-------|-------|

The 8 well cut PCR plate is marked with the Lot No. '6F3' in silver/gray ink.

Well No. 1 is marked with the Lot No. '6F3'.

Well 3 – Negative Control (NC).

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded. The PCR plates are heat-sealed with a PCR-compatible foil.

Please note: When removing each 8 well PCR plate, make sure that the remaining plates stay sealed. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

INTERPRETATION

The interpretation of DRB4*01:03:01:02N SSP subtypings will be influenced by most other DRB4 alleles. For further details see Specificity Table.

UNIQUELY IDENTIFIED ALLELES

The DRB4*01:03:01:02N allele will give rise to a unique amplification pattern by the primers in the DRB4*01:03:01:02N kit^{1,2}.

¹DRB4 alleles listed on the IMGT/HLA web page 2017-April-13, release 3.28.0, www.ebi.ac.uk/imgt/hla.

²Alleles that have been deleted from or renamed in the official WHO HLA Nomenclature up to and including the last IMGT/HLA database release can be retrieved from web page <http://hla.alleles.org/alleles/deleted.html>.

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Lot-specific information
SPECIFICITY TABLE

DRB4*01:03:01:02N SSP subtyping

Specificities and sizes of the PCR products of the 2+1 primer mixes used for DRB4*01:03:01:02N SSP subtyping

| Primer Mix | Size of spec. PCR product ¹ | Size of control band ² | Amplified DRB4 alleles ³ |
|----------------|--|-----------------------------------|---|
| 1 | 155 bp | 515 bp | *01:03:01:02N |
| 2 | 245 bp | 430 bp | *01:01:01:01-01:03:01:01, 01:03:01:03-01:04, 01:05 [?] , 01:06-01:51, 02:01N |
| 3 ⁴ | - | - | Negative Control |

¹Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of DRB4*01:03:01:02N SSP typings. Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers. PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings. PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherent feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

²The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

³For several DRB alleles 1st and/or 3rd exon(s) and beyond, as well as intron nucleotide sequences, are not available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. Assumption is made that unknown sequences in these regions are conserved within allelic groups

⁴Primer mix 3 contains a negative control, which will amplify more than 95% of HLA amplicons as well as the amplicons generated by the control primer pairs matching the human growth hormone gene. HLA-specific PCR product sizes range from 75 to 200 base pairs and the PCR product generated by the HGH positive control primer pair is 430 base pairs.

‘?’, the nucleotide sequence of the primer matching region is not known.

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PRIMER SPECIFICATION

| Well No. | 1 | 2 |
|---------------------------|----------------------|--------------------------------|
| Length of spec. | 155 | 245 |
| PCR product | | |
| Length of int. | 515 | 430 |
| pos. control ¹ | | |
| 5'-primer(s) ² | 5(101) ⁴ | 1 st I ⁵ |
| | 5'-CAA ^{3'} | 5'-ggg ^{3'} |
| 3'-primer(s) ³ | 42(213) | 1 st I ⁶ |
| | 5'-TCA ^{3'} | 5'-TgC ^{3'} |
| Well No. | 1 | 2 |

¹The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases

²The nucleotide position matching the specificity-determining 3'-end of the primer is given. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

³The nucleotide position matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

⁴Matching the sequence of the 3'-end of the 1st intron.

⁵Matching sequences within the 1st intron.

⁶Matching sequences from the 3'-end of the 1st intron into the 5'-end of the 2nd exon.

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| CELL LINE VALIDATION SHEET | | | | | | |
|--|------|------------|---------|--------|------------|-----------|
| DRB4*01:03:01:02N SSP kit ² | | | | | | |
| | | | | | Well | |
| | | | | | 1 | 2 |
| | | | | | Prod. No.: | |
| | | | | | 201784401 | 201784402 |
| IHC cell line ¹ | | | DRB4 | | | |
| 1 | 9001 | SA | | | - | - |
| 2 | 9280 | LK707 | *01:03 | | - | + |
| 3 | 9011 | E4181324 | | | - | - |
| 4 | 9275 | GU373 | | | - | - |
| 5 | 9009 | KAS011 | | | - | - |
| 6 | 9353 | SM | *01:03 | | - | + |
| 7 | 9020 | QBL | | | - | - |
| 8 | 9025 | DEU | *01:01 | | - | + |
| 9 | 9026 | YAR | *01:03 | | - | + |
| 10 | 9107 | LKT3 | *01:03 | | - | + |
| 11 | 9051 | PITOUT | *01:01 | | - | + |
| 12 | 9052 | DBB | *01:03N | | + | - |
| 13 | 9004 | JESTHOM | | | - | - |
| 14 | 9071 | OLGA | | | - | - |
| 15 | 9075 | DKB | *01:03 | | - | + |
| 16 | 9037 | SWEIG007 | | | - | - |
| 17 | 9282 | CTM3953540 | | | - | - |
| 18 | 9257 | 32367 | *01:01 | | - | + |
| 19 | 9038 | BM16 | | | - | - |
| 20 | 9059 | SLE005 | | | - | - |
| 21 | 9064 | AMALA | | | - | - |
| 22 | 9056 | KOSE | | | - | - |
| 23 | 9124 | IHL | | | - | - |
| 24 | 9035 | JBUSH | | | - | - |
| 25 | 9049 | IBW9 | *01:01 | | - | + |
| 26 | 9285 | WT49 | | | - | - |
| 27 | 9191 | CH1007 | *01:03 | | - | + |
| 28 | 9320 | BEL5GB | *01:01 | | - | + |
| 29 | 9050 | MOU | *01:01 | | - | + |
| 30 | 9021 | RSH | | | - | - |
| 31 | 9019 | DUCAF | | | - | - |
| 32 | 9297 | HAG | | | - | - |
| 33 | 9098 | MT14B | *01:03 | | - | + |
| 34 | 9104 | DHIF | | | - | - |
| 35 | 9302 | SSTO | *01:03 | | - | + |
| 36 | 9024 | KT17 | *01:03 | | - | + |
| 37 | 9065 | HHKB | | | - | - |
| 38 | 9099 | LZL | | | - | - |
| 39 | 9315 | CML | *01:02 | | - | + |
| 40 | 9134 | WHONP199 | *01:03 | | - | + |
| 41 | 9055 | H0301 | | | - | - |
| 42 | 9066 | TAB089 | | | - | - |
| 43 | 9076 | T7526 | *01:03 | | - | + |
| 44 | 9057 | TEM | | | - | - |
| 45 | 9239 | SHJO | *01:01 | *01:03 | - | + |
| 46 | 9013 | SCHU | | | - | - |
| 47 | 9045 | TUBO | | | - | - |
| 48 | 9303 | TER-ND | | | - | - |

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¹The provided cell line HLA specificities are retrieved from the <http://www.ihwg.org/hla> web site. The specificity of an individual cell line may thus be subject to change.

²The specificity of each primer solution in the kit has been tested against 48 well characterized cell line DNAs and where applicable, additional cell line DNAs.

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ADDRESSES:

Manufacturer:

Olerup SSP AB, Franzengatan 5, SE-112 51 Stockholm, Sweden.

Tel: +46-8-717 88 27

Fax: +46-8-717 88 18

E-mail: olerup-se@caredx.com

Web page: <http://www.olerup.com>

Distributed by:

Olerup GmbH, Löwengasse 47 / 6, AT-1030 Vienna, Austria.

Tel: +43-1-710 15 00

Fax: +43-1-710 15 00 10

E-mail: olerup-at@caredx.com

Web page: <http://www.olerup.com>

Olerup Inc., 901 S. Bolmar St., Suite R, West Chester, PA 19382

Tel: 1-877-OLERUP1

Fax: 610-344-7989

E-mail: olerup-us@caredx.com

Web page: <http://www.olerup.com>

For information on *Olerup* distributors worldwide, contact **Olerup GmbH**.